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INTERFERENCES IN NITROGEN DETERMINATION METHODS Part 3. Determination of Organic Nitrogen in The Presence of Nitrate KERANCUAN DALAM BEBERAPA METODE PENETAPAN NITROGEN Bagian 3. Penetapan Organik Nitrogen Pada Sampel Yang Mengandung Nitrate

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Summary

Dalam bagian ke-3 ini, penelitian diarahkan untuk mengetahui kemungkinan adanya pengaruh reaksi kimia yang tidak diinginkan dalam penetapan kadar nitrogen terhadap sampel yang mengandung organik N dan nitrate (NO₃), Penetapan NO₃ dilakukan dengan metode MgO, auto-analyzer dan Devarda. Penetapan organik N dilakukan dengan metode Kjeldahi dan total N dengan metode persulfat. Metode tersebut dilakukan untuk mengetahui perolehan kembali nitrogen dari beberapa sampel yang merupakan campuran senyawa organik N dengan NO₃ yang telah diketahui kadar nitrogennya.

Hasil penelitian menunjukkan perolehan kembali nitrogen yang lebih kecil atau lebih besar dari kandungan nitrogen yang sebenarnya. Hal tersebut menunjukkan kecenderungan kuat adanya pengaruh reaksi kimia yang tidak diharopkan. Perolehan kembali NO3 dengan metode Devarda sangat rendah, hal tersebut disebabkan oleh beberapa kemungkinan yaitu menurunnya kapasitas reduksi NO3 menjadi NO2 dan kemudian menjadi NH\$\pi\$ oleh karena pengaruh Cu\$\frac{1}{2}\$, PO\$\frac{1}{2}\$ atau Mg\$_2\$^+ yang digunakan dalam prosedur analisis tersebut. Selain itu diduga kehilangan NO3 sudah terjadi sebelum analisis dilakukan, yaitu oleh karena reaksi NO3 dengan senyawa organik menjadi N2 gas dan dalam kasus tertentu menjadi organik N. Analisis NO3 dengan metode auto-analyzer memberikan perolehan kembali NO3 yang cukup tingsi.

Metode Kjeldahl memberikan perolehan kembali nitrogen yang rendah untuk senyawa yang mengandung amine dan sangat rendah untuk senyawa azo. Hal tersebut disebabkan karakteristik yang lebih dengan adanya NO₃ serta kondisi destilasi yang tinggi (400°C). Metode persulfat relati sangat tinggi dalam perolehan kembali nitrogen dibandingkan dengan metode Kjeldahl, namun dalam beberapa kasus memungkinkan perolehan kembali yang terlempau tinggi.

Reaksi nitrasi atau reaksi nitrat dengan phenol/naphtol terlihat jelas dengan perubahan status NO₃ menjadi organik N pada penetapan nitrogen dengan metode Kjeldahl dan persulfat untuk senyawa campuran α -naphtol dan 1-4-naphto-quinone dengan NO₃.

I. INTRODUCTION

The end product of the aerobic nitrification process, nitrate, is present in quite large amount in environmental samples and is linked with the problem of the eutrophication of surface water. It is known that in soils containing a lot of organic matter significant amount of NO2 is withdrawn from the nitrification process (Boudot and Chone, 1985). At a much lesser extent, this is also the case for normal agricultural soils (Azhar et al, 1989). It is not known whether a similar reaction might take place between NO3 and typical constituents of soil organic matter. There are, however, indications pointing to the possibility of such a reaction: it is well established that organic compounds often interfere with colorimetric methods for the determination of NO; (Norwitz, 1979), while it has been reported that nitrate was fixed by hydrolyzed lignin in a Chernozem soil.

In this paper the reactions between NO, and organic compounds during incubation and analysis were investigated and the reliability of commonly used laboratory methods for the determination of N for samples containing those two components was assessed.

II. MATERIALS AND METHODS

A. Methods of nitrogen determination

Solution containing both organic compounds and NO₃ were subjected to five N-determination procedures. Mineral N was determined by two steam distillation methods, i.e. the MgO and the Devarda method and one automated colorimetric determination, in which nitrate was first reduced to nitrate on a Cd-column (Norwitz,

1979). Two acid digestion methods were used for the determination of resp. organic and total N, i.e. a macro-Kjeldahl and the persulphate method.

B. Reaction between NO; and organic compounds

Solutions buffered at pH 7.0 and pH 5.5 were prepared KNO₃—N was added to a level of 50 mg l⁻¹. The solutions were shaken for three days on a rotary shaker, where after N was determined in threefold.

The % recovery of N as well as the standard deviations were calculated. For mineral N, the recovery was expressed as a % of the NO₃—N added, for Kjeldahl N as a % of both total and organic N and persulphate N as a % of total N.

III. RESULTS AND DISCUSSION

Table 1 gives the averages and the standard deviations of the % N- recovery for the various methods.

Only in two cases, i.e. for adenine (pH 5,5) and aminophenol (pH 7.0), a significant amount of NHI-N appeared have been formed. For the reference compounds as such, no NH4 + was recovered upon Mg0 distillation. Therefore, this phenomenon cannot be contributed to the alkali-labile character of these compounds. The results of the analysis of NO₃-N by the different methods are realigned in Fig. 1 and 2. For the auto-analyzer method, the recovery of NO₃-N is ranged from 70% to 100% and no significant differences could be noticed between the two pH values. Diaminonaphthalene seemed to be an exception to this pattern. The recovery of NO-N is higher for the auto-analyzer method in all cases but one, i.e. diaminonaphthalene and NO3-.

The recovery of NO₃ with the Devarda method in pure KNO₃ solution was over 90%. In a buffered solution containing both NO₃ and organic compound, NO₃—N recovery by the Devarda method was much lower.

In all cases, the recovery was much lower for samples incubated at pH 5.5 than for samples incubated at pH 7.0. One factor explaining the lower N-recovery as compared to the reference may well be the PO₃³— content of the buffer solution. It has been reported that the reduction of NO₃ to NO₂- by a Cd-Cu reduction column was 40% lower in the presence of 25 uM PO₃³- (Olson, 1969). All though in the auto-analyzer method, NO₃ is reduced on a Cd-column, the samples were 100 times diluted. Hence, the actual

Table 1. Percentage recovery of nitrogen for the various methods.

Tabel 1. Persentase perolehan kembali nitrogen untuk metode yang berbeda.

Com- pounds (Senyawa)		%'N recovered Method					
		Mg0	AA	De varda	Kjelda N-org	hl N-tot	Per- sulphate
Adenine + NO ₃	7.0	0.6 0.1	69.7 1.3	26.3 0.9	96.8 1.4	88.3 1.3	68.2* 2.2**
	5.5	26.4 5.5	72.9 0.4	10.1 2.5	25.2 4.5	23.1 4.1	76.8 2.0
Alanine + NO3	7.0	3.2 0.5	90.8 0.5	78.4 5.6	94.4 1.1	71.6 0.9	118.6 1.1
	5.5	1.1 0.8	90.2 0.7	22.1 5.0	94.1 4.5	71.3 0.2	104.1 1.6
Amino phenol + NO3	7.0	23.6 1.2	73.9 0.3	80.9 4.0	80.8 11.2	58.1 8.0	94.2 1.7
	5.5	5.0 0.9	77.2 1.0	20.7 6.9	94.1 1.6	67.6 1.1	90.1 1.8
Benzo- triazole + NO ₃	7.0	0.0 0.0	96.6 1.0	71.0 0.8	88.8 0.2	25.9 0.2	45.6 1.3
	5.5	0.0	95.0 0.6	22.9 0.7	33.9 0.4	26.4 0.3	36.8 1.8
Diamino- naphtha-	7.0	0.0 0.0	48.2 34.2	60.4 4.8	76.8 1.3	59.5 1.0	108.7 11.4
lene + NO ₃	5.5	0.0 0.0	83.6 0.9	8.7 \$.5	41.8 4.3	32.2 3.4	93.8 3.1
Glucos- amine + NO ₃	7.0	0.0 0.0	95.6 0.3	68.9 0.5	104.5 9.4	58.4 5.3	100.9 1.6
	5.5	0.0 0.0	94.8 0.7	25.9 4.8	102.7 2.4	57.4 1.3	96.9 1.7
Naphthol + NO ₃	7.0	0.1 0.1	102.1 0.6	56.6 13.9	<u>-</u>	24.9 4.7	105.3 2.3
	5.5	0.0	102.1 2.2	13.2 2.8	- , -	100.6 18.9	102.9 4.3
Naphtha- quinone + NO ₃	7.0	0.0 0.0	75.2 2.1	19.5 2.1	<u> </u>	9.3 2.5	75.5 8.0
	5.5	0.0 0.0	75.9 1.6	7.0 0.5	-	16.5 2.8	42.6 1.4

Remarks (keterangan): *) Averages (Rata-rata)

**) Standard deviations
(Simpangan baku)

concentration of PO₄³ in the samples was far below this value. For the Devarda method, the samples were undiluted. Since the Devarda alloy consists of 50% Cu, PO₄³ might have interfered with the reduction of NO₃, Mg₂⁺ also has been reported to hamper the reduction of NO₃ during the Devarda procedure (Chausen et al., 1980). It was hypothesized that in acid solution. Mg0

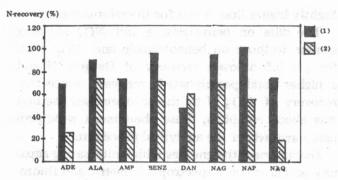


Figure 1. Sample mixtures of organic N and NO₃ kept at pH 7.0 Recovery of NO₃ by two different methods.

Gambar 1. Sampel yang mengandung organik N dan NO₃-pada pH 7,0 Perolehan kembali NO₃ dengan menggunakan dua metode yang berbeda.

(1): Auto-analyzer(2): Devarde

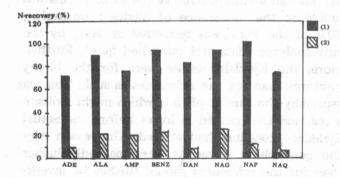


Figure 2. Sample mixtures of organic N and NO₃ kept at pH 5.5 Recovery of NO₃ by different methods.

Gambar 2. Sampel yang mengandung organik N dan NO3 pada pH 5,5 Perolehan kembali NO3 dengan menggunakan dua metode yang berbeda.

1): Auto-analyzer

2): Devarda

was partially dissolved, providing a quantity of Mg_2^+ in the solution, compromising the Devarda reduction alloy. Mg0 was added prior to the Devarda treatment, it was therefore possible that for the pH 5.5 buffer, the amount of Mg0 added was not enough to increase the pH sufficiently to prevent Mg0 dissolution.

The influence of the presence of NO₃ on the recovery of total and organic N in illustrated by Fig. 3 and 4. Fig. 3 gives the Kjeldahl-N recovery rates for organic compounds in the presence of NO₃- as well as for organic compounds as such.

Three cases can be distinguished: (1). No marked influence of the presence of NO₃: alanine, aminophenol and benzotriazole; (2). Loss of organic N: adenine and diaminonaphthalene; (3). Increase of organic N: naphthol and naphthoquinone. The data on total Kjeldahl (Table 1) show that in one case, i.e. naphthol at pH 5.5, there was a complete reaction between NO₃ and the organic compound: 100% of the mineral N was recovered as Kjeldahl N. When there was an influence of the NO₃ presence, Fig 3 shows that it was most likely to occur in samples incubated at pH 5.5.

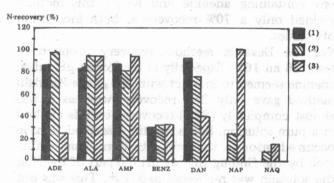


Figure 3. Determination of organic N by the Kjeldahl N method.

Gambar 3. Penetapan organik N dengan menggunakan metode Kjeldahl.

(1): Organic compound as such

(2): Organic compound Kept with NO₃ at pH 7.0

(3): Organic compound kept with NO₃ at pH 5.5

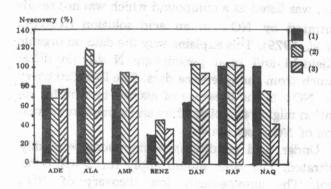


Figure 4. Determinations of total N by the persulphate method.

Gambar 4. Penetapan total N dengan menggunakan metode persulphate

(1): Organic compound as such

(2): Organic compound kept with NO 3 at pH 7.0

(3): Organic compound kept with NO 3 at pH 5.5

Fig. 4 shows the persulphate N-recovery of the organic compounds in the presence NO₃, the combined N-recovery for NO₃ and the organic compounds as determined separately. There was a clear loss of N for naphthoquinone, a clear increase of N for diaminonaphthalene and minor or no changes for the other compounds. The recovery of total N was deficient for adenine, benzotriazole, and naphthoquinone while it was about complete for the cases.

In the presence of adenine, NO₂- was adequately recovered by the auto-analyzer method. In solutions containing adenine and NO₃, this method yielded only a 70% recovery at both incubation

pH values.

For the Devarda method, recovery dropped to resp. 26 an 10%. Especially at the lowest pH value, adenine seemed to interact with NO₃: the Kjeldahl method gave only 25% recovery, whereas N was almost completely (86%) recovered by this method in a pure solution. Some reactions seemed to have occurred prior to the acid digestion. This confirmed by the finding that at pH 5.5 part of the N in the solution was recovered as NH 4+. This was not merely an artifact of the Mg0-method; in a separate set of analyses, NH 4 was also recovered by the auto-analyzer using indophenol blue method (data not published here). The reason for this phenomenon is not clear.

Except for the Devarda determinations, alanine nor NO₅ seemed to affect to a great extent the resp. N determinations. Another ammo acid., L-asparagine, was listed as a compound which was not readly nitrated by NO₅ in an acid solution (Nortwiz et al, 1979). This explains why the data on organic Kjeldahl and total persulphate N do not differ much from the reference data. The lower recovery of NO₅ in the presence of alanine in a buffer solution might be explained by an incomplete reduction of NO₅ (see above).

Under actid conditions, aminophenol is readily nitrated. The same should apply for 2-aminophenol. The unexpectedly low recovery of NO 3 (only 70%) by the auto-analyzer method could be due to this phenomenon. The fact the Kjeldahl recovery rates were not very much different from the data for the reference compound does not contradict this: the resulting product of an acid nitration is likely to be a nitrocompound, which is only partly recovered by the Kjeldahl method. Though for the persulphate method, recovery of amino-N and nitro-N is more or less the same, total N recovery by the persulphate method was

slightly higher than it was for the references.

The data on benzotriazole and NO₃ are very similar to those on benzotriazole and NO₂: i.e. for pH 5.5 a lower recovery of Devarda N and a higher total persulphate-N recovery. Since the recovery of NO₃ N by the auto-analyzer method was about complete, these phenomena were probably artifacts of the analytical procedure.

Diaminonaphthalene has a high nitrosating capacity and is acid as a spectrophotometric and fluorometric reagent for the determination of NO₂. Even though there was no interference of NO₃ on this method, it cannot be excluded that nitrosation takes place on another site of the molecule. Diaminonaphthalene indeed seemed to react with NO: the recovery of NO: by the auto-analyzer method at pH 7.0 was rather poor (50%) on average, but had a large variability (34% S.D.). This was linked to the occurence of nitrite: on average 20% of the NO3. was recovered as NO2 by the ruto-analyzer (data not published here). Furthermore, the Kjeldahl results were for the binary mixtures than for the references as such. This was especially the case at pH 5.5, which might indicate a reaction that caused N losses before the actual Kjeldahl procedure had started. The reason why the persulphate N results were markedly higher than for the references should further be investigated.

For N-acetylglucosamine, there was only a difference between the pH values for the Devarda method. Since no determination on NAG as such were carried out, the impact of the presence of the NO₃ is difficult to assess.

Phenolic compounds are easily nitrated under acidic conditions (Norwitz, 1979). This reaction is actually used for the determination of NO₃. The end products of the nitration of naphthol are nitro- and dinitronaphtols. As the auto-analyzer data indicate, the nitration at pH 7.0 and 5.5 was either reversible or non-existent. The Kjeldahl procedure, however, recovered organic N, which was not initially present. At pH 5.5, even all NO₃ N initially present was converted to organic N. Nitration seemed to have taken place during the acid digestion. As only a minor fraction of Nitro-N was recovered by the Kjeldahl method, it was unlikely that nitro compounds are the sole end product of this acid digestion catalyzed nitration.

Naphthoquinone too became nitrated, probably

also at neutral pH (only 75% recovery of NO₃ N for the auto-analyzer method) and 10 to 15% of the NO₃ N initially present was recovered as Kjeldahl -N. The reaction was probably linked to N-volatilization since there was a net-deficit of total persulphate N, which was more obvious at the lower pH.

IV. CONCLUSIONS

Determination of nitrogen in samples containing NO₃ and organic compounds did not always give accurate results. The compounds reacted and interfered with one another before and during the N determination procedures. This caused under or over estimation of the actual amount of N present in the samples. The following cases could be distinguished:

 Compounds containing an amino group were liable to decompose during the Mg0 treatment when NO₃- as well was present in the samples;

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- The auto-analyzer method gave the best recovery results for NO₃, the recovery with the Devarda method was much lower;
- Organic N in phenolic compounds was excessively recovered by the Kjeldahl method. Nitration is likely to have occured during the digestion.

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